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Antimicrobial Activity of Lidocaine, Bupivacaine, Mepivacaine and Ropivacaine on *Staphylococcus epidermidis*, *Staphylococcus aureus* and

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Abstract:

Introduction:

Bacillus subtilis

Various studies have shown a possible antimicrobial activity of different local anaesthetics, which may affect the results of microbial assessment of biopsies. The purpose of this study was to test the antimicrobial activity of different commonly used anaesthetic agents on *Staphylococcus epidermidis, Staphylococcus aureus* and *Bacillus subtitles* to reproduce data and to compare the findings.

Methods:

Local anaesthetics tested were commercially available solutions of lidocaine (*Xyloneural*[®], *Xylanaest pur.*[®]), bupivacaine (*Bucain*[®]), mepivacaine (*Mecain*[®]) and ropivacaine (*Naropin*[®], *Ropinaest*[®]).2%, 1%, 0.5%, 0.25% and 0.1% (20, 10, 5, 2.5, 1 mg/ml) dilutions of these local anaesthetics were prepared with sterile 0.9% saline. Bacteria used in this study were *Staphylococcus epidermidis*, *Staphylococcus aureus* and *Bacillus subtilis*. 10 µl of different local anaesthetic dilution placed on thin wafers were added to Mueller Hinton Agar and cultured. After 24 hours, a zone of inhibition around the wafers was evaluated.

Results:

Local anaesthetics in different concentrations did not show any zone of inhibition on *Staphylococcus epidermidis, Staphylococcus aureus* or *Bacillus subtilis*.

Conclusion:

In summary, neither lidocaine, bupivacaine, mepivacaine nor ropivacaine showed an antibacterial effect on *Staphylococcus* epidermidis, *Staphylococcus aureus* and *Bacillus subtilis*.

Implications:

Due to these findings this local anaesthetics can be used in daily clinical routine to perform pain free diagnostic procedures in which culture specimens are to be obtained. Due to inconsistent results in prior studies, we recommend to use the lowest concentration possible of the local anaesthetic, also to avoid other possible side effects of local agents.

Keywords: Pain, Local anaestetic, Concentration, Antibacterial effect.

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INTRODUCTION

In clinical practice local anaesthetics are used daily to perform pain free diagnostic procedures. However, various studies have shown a possible antimicrobial activity of different local anaesthetics, which may affect results in microbial assessment of biopsies. The purpose of this study was to test the antimicrobial activity of different commonly used anaesthetic agents on *Staphylococcus epidermidis*, *Staphylococcus aureus* and *Bacillus subtilis* to reproduce data and to compare the findings.

METHODS

Local anaesthetics tested were commercially available solutions of lidocaine (*Xyloneural*[®], *Xylanaest pur.*[®]), bupivacaine (*Bucain*[®]), mepivacaine (*Mecain*[®]) and ropivacaine (*Naropin*[®], *Ropinaest*[®]). 2%, 1%, 0.5%, 0.25% and 0.1% (20, 10, 5, 2.5, 1 mg/ml) dilutions of these local anaesthetics were prepared with sterile 0.9% saline. Bacteria used in this study were *Staphylococcus epidermidis*, *Staphylococcus aureus* and *Bacillus subtilis*. The number of bacteria was prepared to achieve 0.5 McFarland standard and plated. 10 µl of different local anaesthetic dilution placed on thin wafers were added to Mueller Hinton Agar and cultured at 35°C. Wafers without local anaesthetics but 0.9% saline and wafers with ampicillin/sulbactam were used as controls. After 24 hours, a zone of inhibition around the wafers was evaluated, which is a wide ring around the wafer with no bacterial growth.

RESULTS

Local anaesthetics in different concentrations did not show any zone of inhibition on *Staphylococcus epidermidis*, *Staphylococcus aureus* or *Bacillus subtilis*, which means that these bacteria have grown although there was a local anaesthetic (Figs. 1 and 2). Due to these findings the tests were repeated with commercially available undiluted local anaesthetics but with lower densities of microorganisms, according to McFarland standard of 0.25, 0.10 and 0.05 and 100 μ l of different local anesthetic solution. After culturing for 24 hours at 35°C there again was no zone of inhibition on tested bacteria and no change in the surrounding bacterial concentration. In summary, 240 combinations of bacterial and local anaesthetics were tested (Table 1).



Fig. (1). Local anaesthetics in different concentrations did not show any zone of inhibition on Bacillus subtilis.

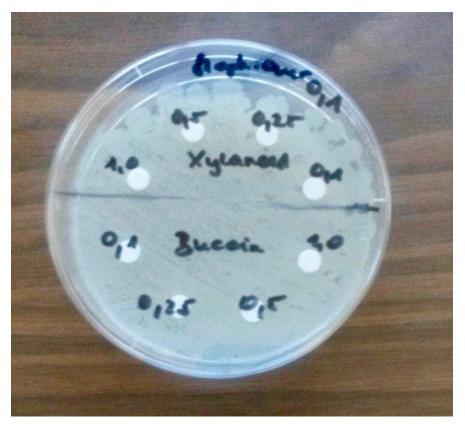


Fig. (2). Local anaesthetics in different concentrations did not show any zone of inhibition on *Staphylococcus aureus*.

| Tested local | | Mc Farland standard | | | | | | | | | | | |
|--------------------|-------|-------------------------------|------|------|------|--------------------------|------|------|----------------------|------|------|------|------|
| anaesthetics | | Staphylococcus epidermidis | | | | Staphylococcus aureus | | | Bacillus subtilis | | | | |
| [| conc. | 0,50 | 0,25 | 0,10 | 0,05 | 0,50 | 0,25 | 0,10 | 0,05 | 0,50 | 0,25 | 0,10 | 0,05 |
| Xyloneural® | 1,0 | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm |
| Xylanaest | 0,5 | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm |
| pur [®] . | 0,25 | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm |
| | 0,1 | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm |
| Bucain® | 1,0 | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm |
| | 0,5 | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm |
| | 0,25 | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm |
| | 0,1 | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm |
| Mecain® | 1,0 | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm |
| | 0,5 | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm |
| | 0,25 | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm |
| | 0,1 | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm |
| Naropin® | 1,0 | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm |
| | 0,5 | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm |
| | 0,25 | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm |
| | 0,1 | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm |
| Ropinaest® | 1,0 | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm |
| | 0,5 | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm |
| | 0,25 | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm |
| | 0,1 | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm | 0 mm |

Table 1. Local anaesthetics / zone of inhibition.

DISCUSSION

In clinical practise local anaesthetics are used daily to perform pain free diagnostic procedures. Considering the literature [1], antimicrobial activity of local anaesthetics could lead to false-negative results in microbial assessment of biopsies. Potential mechanisms of these antimicrobial effects are lysis, increased cell wall permeability and dehydrogenase activity [2]. Various studies conclude that prior to diagnostic biopsies the lowest concentration of local agents should be used or a ring block with additive-free local anaesthetics should be performed [1, 3]. In 2010, *Kose and colleagues* [4] showed that under *in vivo* conditions different local anaesthetics did not have any antimicrobial effects. Also *Aydin et al.* [5] found no antimicrobial properties of ropivacaine but antimicrobial *effectiveness* of lidocaine and prilocain was observed. The antibacterial activity of lidocaine was negative in a study performed by *Berg et al.* [3] but they showed that EMLA[®] has a powerful antibacterial effect. Due to these inconsistent results this present study was conducted to evaluate the antimicrobial effects of different commonly used anesthetic agents on *Staphylococcus epidermidis, Staphylococcus aureus* and *Bacillus subtilis.* Neither lidocaine, bupivacaine, mepivacaine nor ropivacaine showed an antibacterial effect. We feel that our large sample size allows us to make a valid conclusion.

CONCLUSION

In summary, neither lidocaine, bupivacaine, mepivacaine nor ropivacaine showed an antibacterial effect on *Staphylococcus epidermidis*, *Staphylococcus aureus* and *Bacillus subtilis*. Due to these findings this local anaesthetics can be used in daily clinical routine to perform pain free diagnostic procedures in which culture specimens are to be obtained. Due to inconsistant results in prior studies we recommend to use the lowest concentration possible of the local anaesthetic, also to avoid other possible side effects of local agents.

ETHICS APPROVAL AND CONSENT TO PARTICIPATE

Not applicable.

HUMAN AND ANIMAL RIGHTS

No Animals/Humans were used for studies that are base of this research.

CONFLICT OF INTEREST

The authors confirm that this article content has no conflict of interest.

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Declared none.

REFERENCES

- Johnson SM, Saint John BE, Dine AP. Local anesthetics as antimicrobial agents: a review. Surg Infect (Larchmt) 2008; 9(2): 205-13. [http://dx.doi.org/10.1089/sur.2007.036] [PMID: 18426354]
- Leung YW, Rawal BD. Mechanism of action of tetracaine hydrochloride against *Pseudomonas aeruginosa*. J Infect Dis 1977; 136(5): 679-83. [http://dx.doi.org/10.1093/infdis/136.5.679] [PMID: 410897]
- [3] Berg JO, Mössner BK, Skov MN, Lauridsen J, Gottrup F, Kolmos HJ. Antibacterial properties of EMLA and lidocaine in wound tissue biopsies for culturing. Wound Repair Regen 2006; 14(5): 581-5. [http://dx.doi.org/10.1111/j.1743-6109.2006.00157.x] [PMID: 17014670]
- Kose AA, Karabaggli Y, Kiremitci A, Kocman E, Cetin C. Do local anesthetics have antibacterial effect on *Staphylococcus aureus* under *in vivo* conditions? An experimental study. Dermatol Surg 2010; 36(6): 848-52.
 [http://dx.doi.org/10.1111/j.1524-4725.2010.01559.x] [PMID: 20618369]
- [5] Aydin ON, Eyigor M, Aydin N. Antimicrobial activity of ropivacaine and other local anaesthetics. Eur J Anaesthesiol 2001; 18(10): 687-94. [http://dx.doi.org/10.1097/00003643-200110000-00008] [PMID: 11553246]

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